

# Detection of avian influenza

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Reference Laboratory

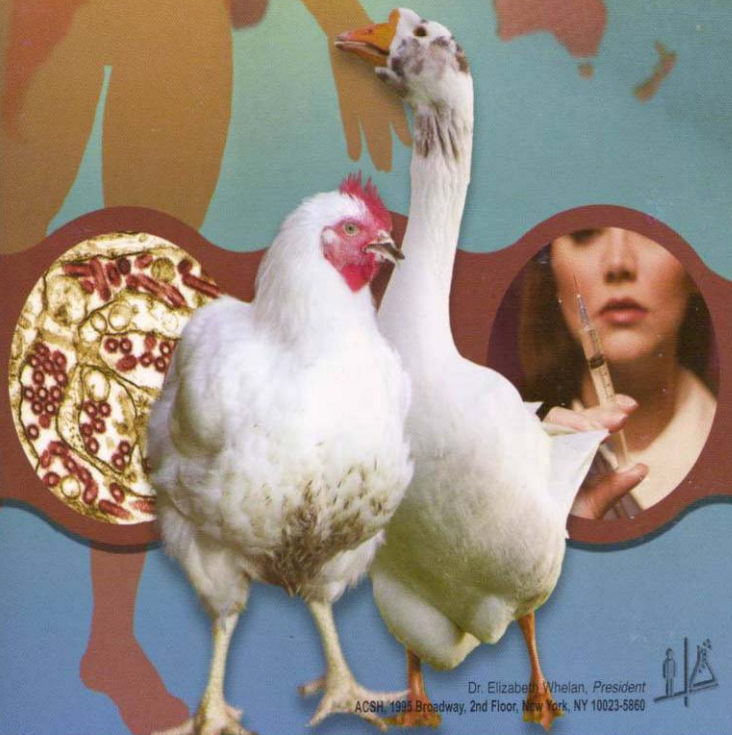
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Institute, CAAS

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# Avian Influenza, or “Bird Flu”: What You Need to Know



Dr. Elizabeth Whelan, President  
ACSH, 1926 Broadway, 2nd Floor, New York, NY 10023-5860



# Avian influenza

- ☞ A contagious disease
- ☞ Domestic poultry flocks are especially vulnerable
- ☞ The disease in birds has two forms: mild or highly pathogenicity (HPAI)

# Common domestic hosts

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<http://www.wwfchina.org/birdgallery/>  
<http://www.wwfchina.org/bbs/guanniao.htm>



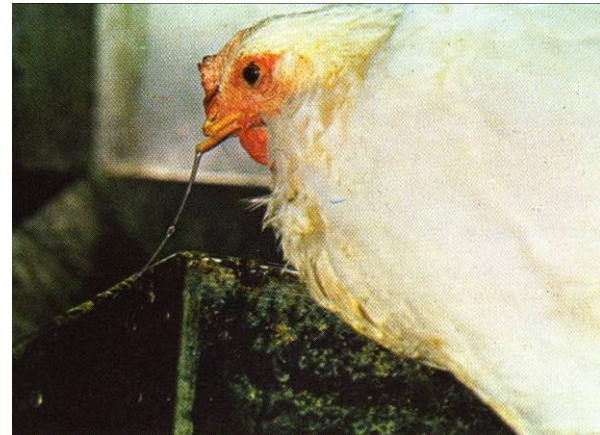
段文科鸟类摄影





# Clinical Manifestations

- Vary greatly depending on age, species, virus virulence, other infections, and production management
- Low Pathogenic AI
  - Depression
  - Respiratory signs
  - Lower productivity
    - egg drop in layers, such as H9N2 infection
  - “Sleepy” chicks



感染减蛋综合征鸡群所产蛋壳质量变差，软皮蛋、无壳蛋

# History of avian influenza

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- First report in Italy in 1878 as Fowl Plague
- Frequent outbreaks up to about 1950
- Confirmed to be caused by influenza A virus in 1955
- First duck isolate (H4N6) in Czechoslovakia in 1956
- First chicken isolate (H5N1) in Scotland in 1959
- First turkey isolate (H6N8) in Canada and H7N3 in England in 1963

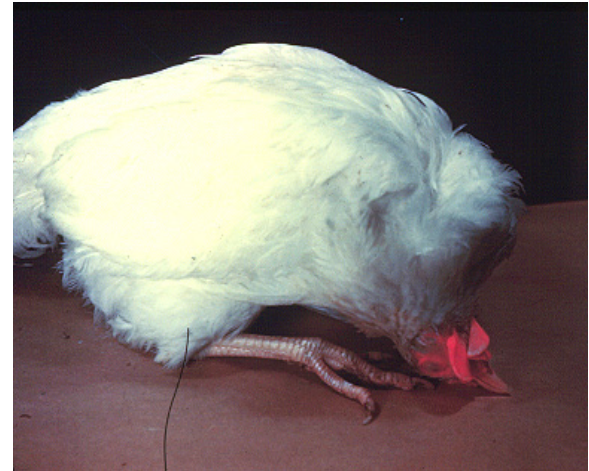
# History of avian influenza

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- Only 21 outbreaks from 1959 to 1997
- Most outbreaks very small and quickly controlled
- A few large widespread epidemics (market dissemination)
- First instance of human infection (conjunctivitis) caused by AIV (H7N7) in England in 1996
- First instance of human's death caused by AIV (H5N1) in Hong Kong in 1997
- Multiple H5 and H7 outbreaks since 1997

# Highly Pathogenic Avian Influenza—Fowl Plague





- Sudden onset high mortality
- Depression, +/- Nervous signs
- Face edema & hemorrhagic lesions



# High Pathogenecity Avian influenza

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How about HPAI in recent years?

-  Since mid-2003, the H5N1virus has caused the largest and most severe outbreaks in poultry on record.
-  In 2005, H5N1 HPAI outbreak in migratory birds in Qinghai lake in western China, It was the first case in the history of avian influenza.
-  Now, the H5N1 HPAIVs have spread all over the world through the migration of wild birds
-  Many human cases



# History of HPAI in domestic poultry

1959-Scotland, H5N1  
1961-S. Africa, H5N3  
1963-England, H7N3  
1966-Canada, H5N9  
1975-Australia, H7N7  
1979-England, H7N7  
1983-84 - USA, H5N2  
1983-Ireland, H5N8  
1985-Australia, H7N7  
1991-England, H5N1  
1992-Australia, H7N3  
1994-Australia, H7N3  
1994-95-Mexico, H5N2

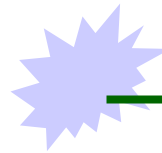
Red  
represents  
shift from  
LPAI to HPAI

1995, 2001 & 2004 -Pakistan,  
H7N3

1997-Australia, H7N4

1997-Italy, H5N2

1996-2007 - Asia, Euro,  
Africa ,H5N1



Largest epizootic in 50 yrs

1999-2000 - Italy, H7N1

2002 - Chile, H7N3

2003 - Netherlands, H7N7

2004 - USA, H5N2

2004 - Canada, H7N3

2004 - S. Africa, H5N2

2005 - N. Korea, H7N?

# The Hong Kong 1997 outbreak

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- Substantial outbreak of H5N1 in domestic poultry
- First began to show mammalian virulence
- 18 human cases, 6 deaths
- Pigs have remained insignificant so far in the epidemiology of H5N1 virus
- The HK97 virus was eradicated by eliminating domestic poultry, has never been detected subsequently

# Evolution 1997-2003

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- H5N1 viruses were isolated on only a few occasions
- Hong Kong had intermittent outbreaks in poultry and a cluster of human cases
- Isolated from two geese in a market in Vietnam in 2001
- Sudden explosive epidemic in Asia and caused much more human cases from 2003

# HPAI diagnosis

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 Clinical

 Post Mortem Lesions

 Virology (Antigen) : usually by virus isolation

Presence of virus confirmed by

AGID

ELISA

RT-PCR

 Serology (Antibody) may be helpful



# Clinical Signs

- ☞ Incubation period: 3-14 days
- ☞ Depression, anorexia, ruffled feathers
- ☞ Neurological signs
- ☞ Conjunctivitis and respiratory signs
- ☞ Drop in egg production
- ☞ Birds found dead
- ☞ Mortality almost 100%



# Clinical Signs

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- ☼ Facial edema
- ☼ cyanotic combs and wattles
- ☼ Petechial hemorrhages on the leg



# Post Mortem Lesions

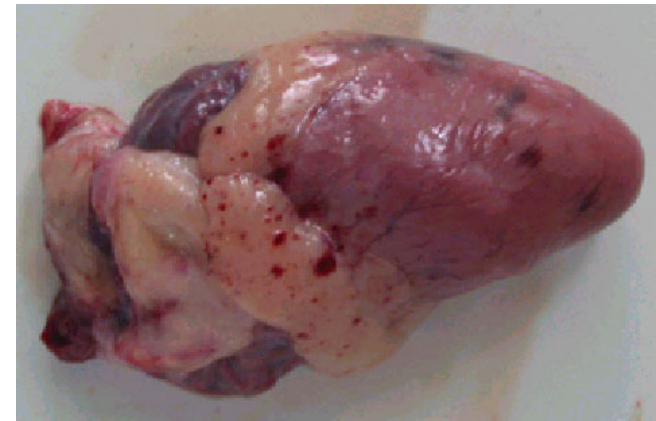
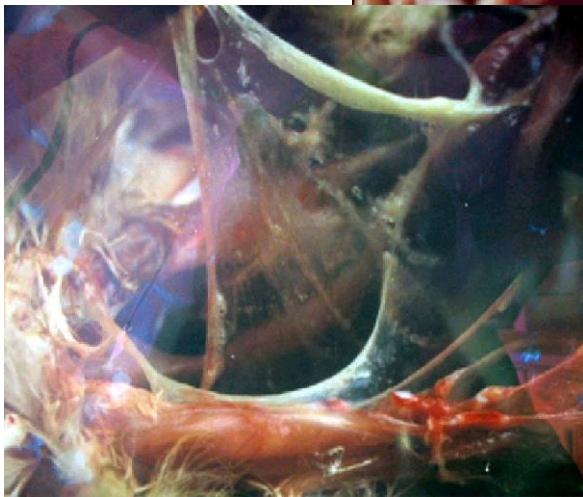
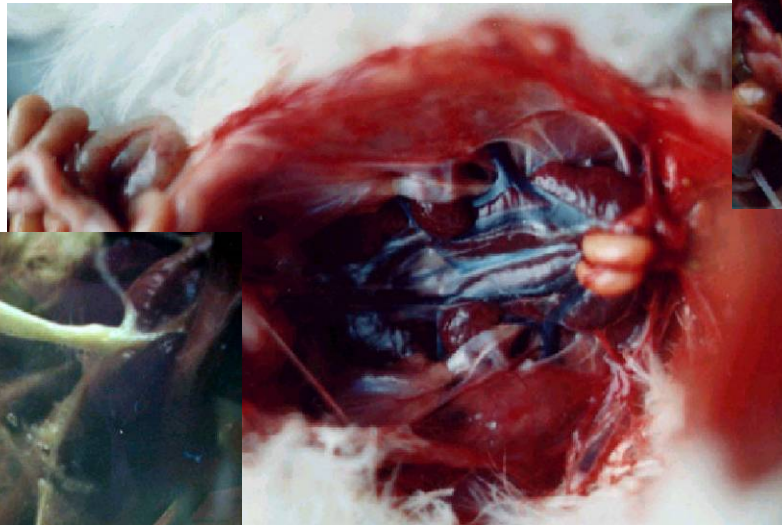
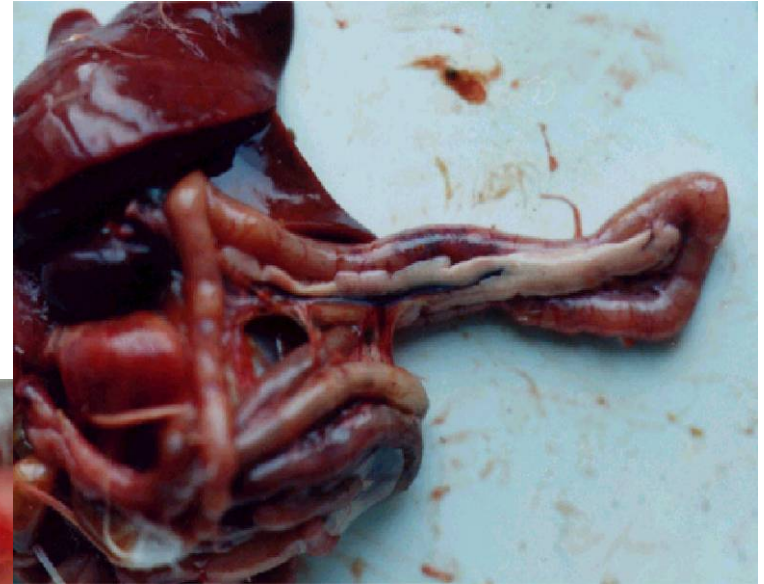
- Lesions may be absent with sudden death
- Severe hemorrhage on tissues
- Subcutaneous edema of head and neck area
- Dehydration





# Post Mortem Lesions

- ☞ Nasal and oral cavity discharge
- ☞ Petechiae on serosal surfaces
- ☞ Kidneys severely congested
- ☞ Intestine hemorrhage





# Post Mortem Lesions

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HPAIVs always can cross the barrier of Blood and Brain, infect brain, then lead to death

Brain is the first and foremost tissue for isolate the HPAIV



# Laboratory Techniques

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- Collection of specimens
- Isolation of influenza virus

# Collection of specimens

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**Specimen** is a significant factor for virus diagnosis

- the quality of the specimen
- the conditions for transport and storage

# Representative Requirements of the Samples

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- 30 Samples/poultry farm
- 30 Samples/live live market
- 20 Samples/habitat
- 20 Samples/swine farm





# The quality of the specimen

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- Should be taken during the first 3 days
- Respiratory & intestinal tract: Avian
- Respiratory tract:  
Mammals: human, pig, horse
- Upper respiratory tract: nasal swab  
throat swab  
tracheal swab
- Intestinal tract: cloaca swab

# Sampling

Samples: a critical element for diagnosis

- 🌐 Collected in day 3 should be best
- 🌐 Swabs: cloacal and laryngeal
- 🌐 Tissues: **Brain**, spleen, kidney, pancreas, lung, trachean
- 🌐 Keep them respectively

Kept in special box with ice



# Sampling

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## Samples: a critical element for diagnosis

- ☞ Before collecting or sending any samples, the proper authorities should be contacted
- ☞ Samples should only be sent under secure conditions and to authorized laboratories to prevent the spread of the disease
- ☞ HPAI samples may be zoonotic
- ☞ Protect yourself when contact HPAI samples at any time

# The quality of the specimen

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## ■ Avian species

live: cloacal, tracheal swab, fecal sample,

dead: together with internal organs

## ■ Mammals

live: tracheal swab/trachea

dead: together with bronchoalveolar lavage,  
lung biopsy sample



# Transport and Storage

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- Sample should kept on ice and processed within 1-2h for immunofluorescence staining
- Chilled in an ice pack immediately for virus isolation ASAP
- Kept frozen at or below  $-70^{\circ}\text{C}$   
If cannot be processed withing 48-72h
- Should not be stored or shipped in dry ice ( $\text{CO}_2$ ), unless be sealed or doubled plastic bagged
- Repeated freezing and thawing must be avoided

# Transport and Storage

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- **Transport medium:** should be supplemented with antibiotics, antimycotics, protein
- **Sterilize, distribution, in ice or in liquid nitrogen**
- **Commonly:** Glycerol medium  
Medium 199
- **Commercially:** Hanks balanced salt solution  
phosphate buffered saline  
cell culture medium, etc.

# Take samples

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- **Nasal swab:** inserted into, parallel to, left for, slowly withdrawn, rotating motion
- **Throat swab:** posterior pharynx, swab vigorously
- **Tracheal swab:** live bird, gently  
dead, vigorously
- **Cloacal swab:** insert deeply, swab vigorously

# Take samples

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- **Fecal sample:** freshly deposited wet feces
- **Tissue sample:** frozen immediately without transport medium, then ground
- **Water sample**
- **Sera:** acute phase , within 1 week;  
convalescent phase, 2-4 weeks.

Blood clot, then centrifuged at 2500rpm,  
15 mins, discard RBC

Stored at -20 °C

# Take samples

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- **Essential to record**

  - Type of animal sampled

  - Type of sample

  - Location of sampled

  - Date of sampled

- Samples collected from cages are called environmental since the source animal is usually uncertain

# Isolation of influenza virus

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## Processing clinical material

- swab samples: thaw, add antibiotics, agitate on vortex, leave for 30 mins at room temp
- Tissue samples: grind, make a 10% (g/v) suspension, add antibiotics, centrifuge at 400xg for 10 mins, inoculate supernatant



# Isolation of influenza virus

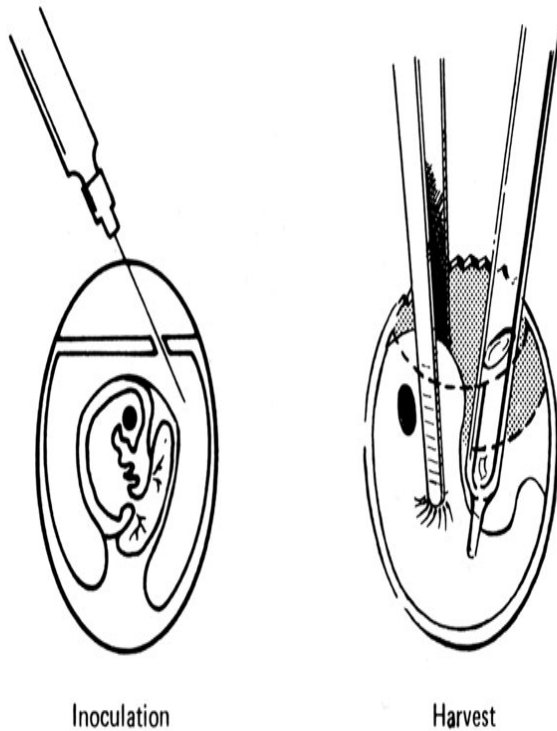
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## Two culture systems

- **Embryonated Egg:** The option of choice for avian influenza viruses, but some of the human or swine influenza virus may grow poorly.
- **Cells:** Used for growing both human and animal influenza viruses. Not suitable for isolation human vaccine candidate
- **Two culture systems combine**

# Isolation from Embryonated Egg

## ALLANTOIC TECHNIQUE



- Canding
- Sterilization
- Inoculation
- Incubation
- Harvesting

# Isolation from Embryonated Egg

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## ■ Canding

discard those are infertile,  
underdeveloped, have cracks, have a  
porous shell

## ■ Sterilization

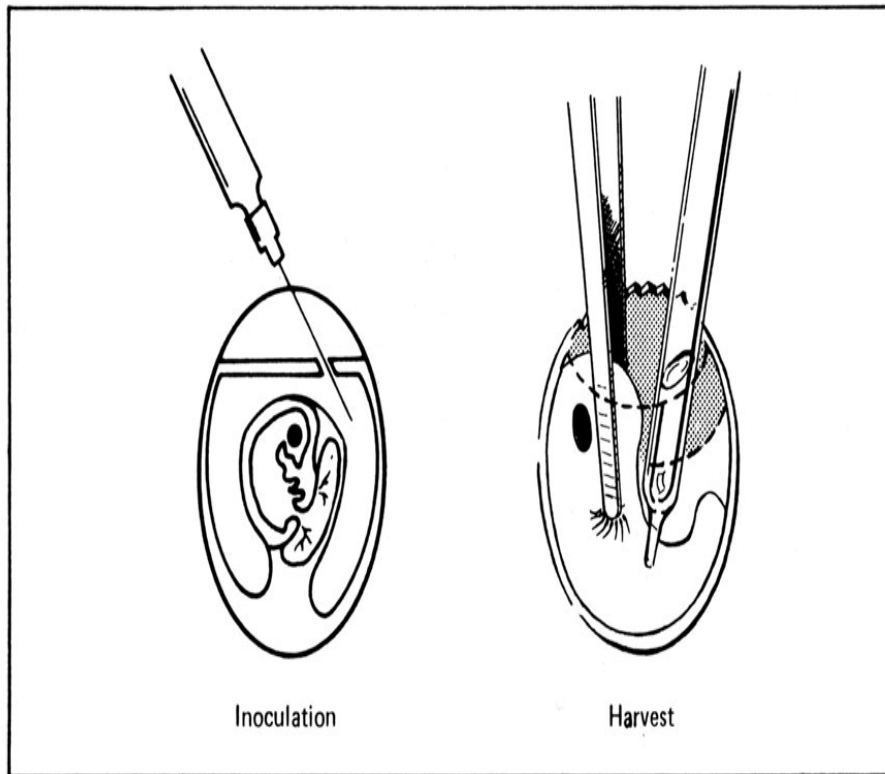
Blunt end up, wipe with 70% ethanol

# Isolation from Embryonated Egg

■ Inoculation: two routes

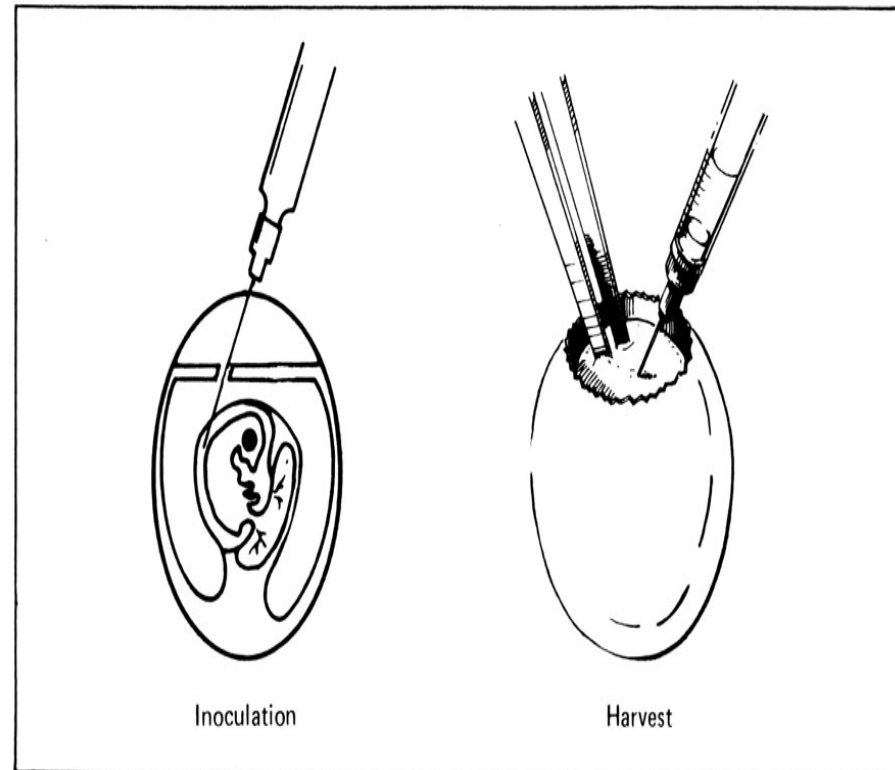
Allantoic cavity

ALLANTOIC TECHNIQUE



Amniotic

AMNIOTIC TECHNIQUE



# Isolation from Embryonated Egg

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## ■ Incubation

Avian influenza virus 35-37 °C

Mammalian influenza virus 35 °C

HPAIV: often dead in 1-2 days

LPAIV: last for 2-4 days

# Isolation of influenza virus

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## ■ Harvest

Chill 4 °C overnight or 4h before harvest

Sterilize

Combine

Centrifuge remove blood cell

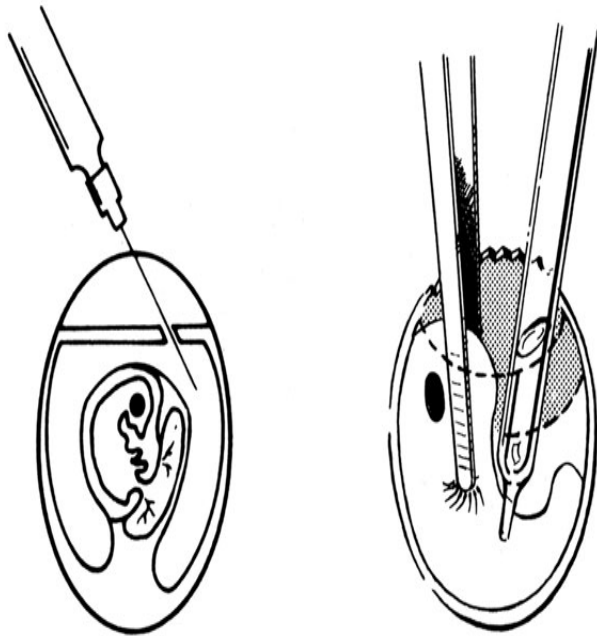
HA test



# Isolation from Embryonated Egg

## ■ Harvest

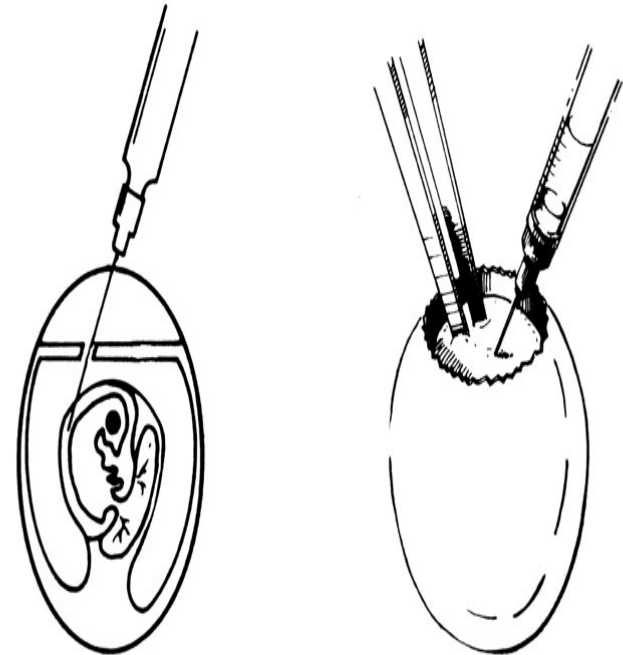
ALLANTOIC TECHNIQUE



Inoculation

Harvest

AMNIOTIC TECHNIQUE



Inoculation

Harvest

# Cell systems

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- MDCK: Madin-Darby Canine Kidney: most common used cell line
- CEF: Chick Embryo Fibroblast; most sensitive for isolation avian isolates

# Cell systems

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- MDCK: Madin-Darby Canine Kidney: most common used cell line
- CEF: Chick Embryo Fibroblast; most sensitive for isolation avian isolates
- GMK: Green Monkey Kidney; most sensitive for human and swine isolates
- Vero: Special licensed acceptable for human vaccine production

# MDCK

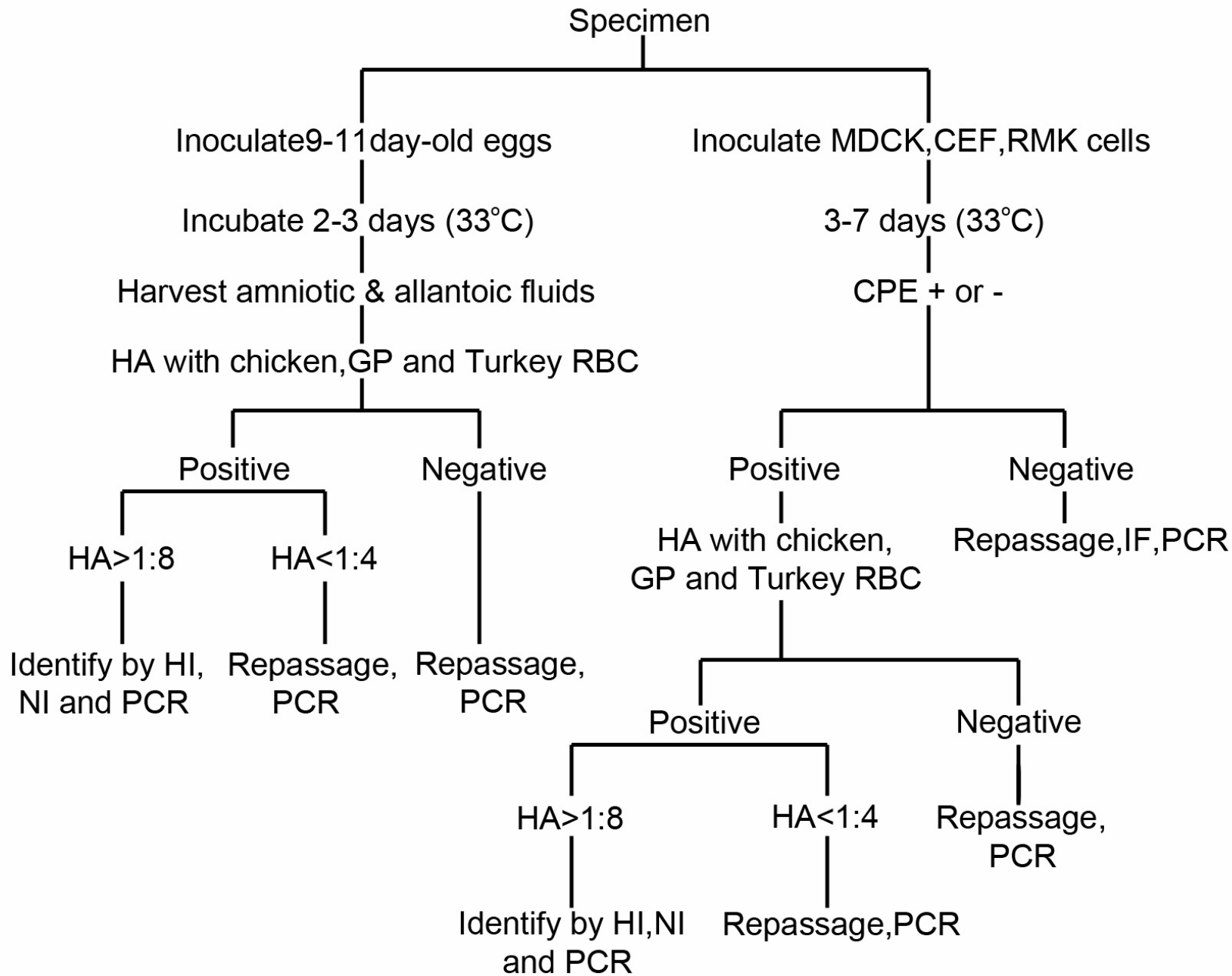
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- After certain passages , the sensitivity of the isolation become low
- Different source of MDCK cells differ on the sensitivity of the isolation
- Virus isolated from MDCK cells more-like the original clinical samples than egg-grown viruses
- No serum in the virus culture medium and don't forget add trypsin to virus culture medium especially for isolation attenuated avian or mammalian virus

# Isolation from MDCK

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- Wash the cells with no serum medium
- Inoculation of the sample
- Absorption: RT/37 °C, 30mins
- Take off the inoculum and add VGM
- Incubation: 37 °C, 3-7 days
- Observation: CPE cytopathic effect
- Harvesting
- Detection: HA, Haemadsorption, IF, PCR
- Repassage





# Precautions

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- **Golden rule1:** Never process clinical specimens for virus isolation and laboratory-adapted influenza strains at the same time
- **Golden rule2:** Never process clinical specimens from humans and from swine or birds in the same laboratory
- **No HA does not mean no virus, before report negative blind passage 2-3 times**

# Storage of influenza virus

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- 4 °C : 1-2 days
- -80 °C : years
- Lyophilised: most stable

# Laboratory Record Keeping

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## ■ Strain designations

The antigenic type of nucleoprotein  
A,B,C

host origin, locus, year subtype

A/Goose/Guangdong/1996(H5N1)

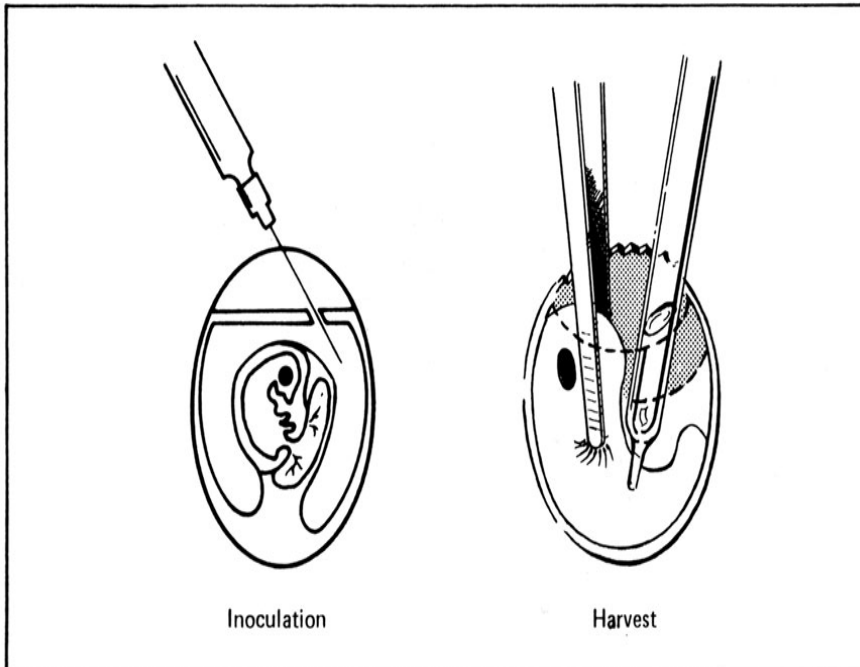
# Diagnosis-Virus isolation

Virus isolation: necessary, useful

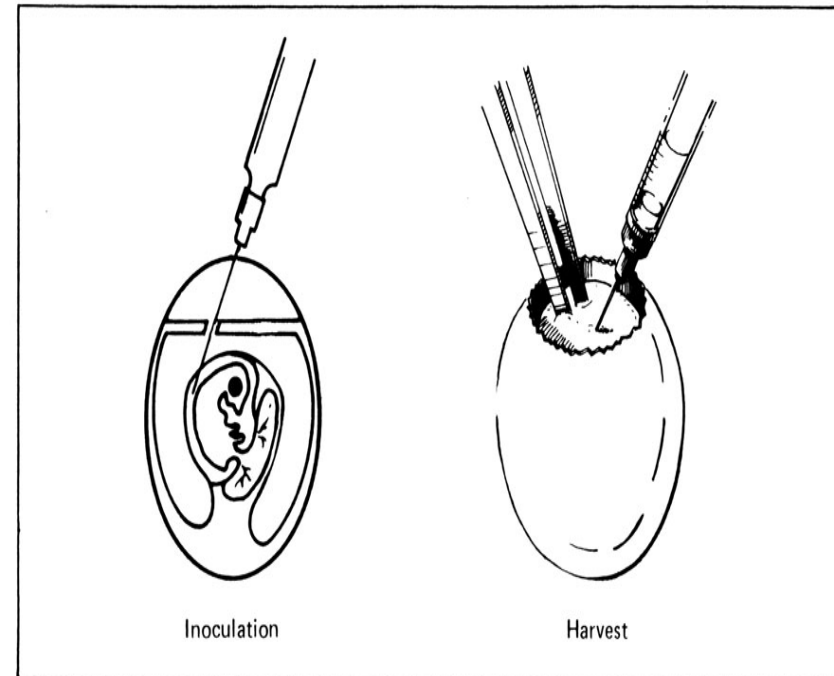
BSL-3 conditions

**Egg** or cell culture

ALLANTOIC TECHNIQUE







AMNIOTIC TECHNIQUE



# Diagnosis-Virus isolation

## Incubation

-  Avian influenza virus: 35 °C -37 °C
-  Mammalian virus: 35 °C
-  HPAIV: dead in 1~2 days
-  LPAIV: last 2~4 days

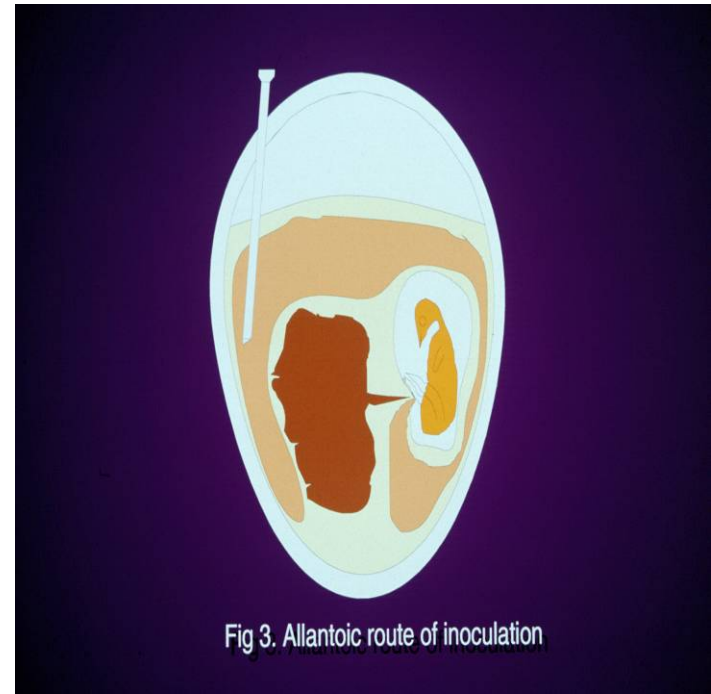


Fig 3. Allantoic route of inoculation

# Diagnosis-Virus isolation

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## Advantages:

- 🌐 Gold rule: the recommended method by WHO,OIE
- 🌐 Accurate: with the highest accuracy
- 🌐 Useful: for the following researches

## Disadvantages:

- 🌐 Time waste: more than one day usually
- 🌐 Laboratory security: BLS-3
- 🌐 Higher quality of sample
- 🌐 The other pathogen should be exclude
- 🌐 Intercross contamination

So serology and some rapid diagnosis technology have been developed



# Diagnosis-serology

Serology methods:

AGP

ELISA

VNT

IFA

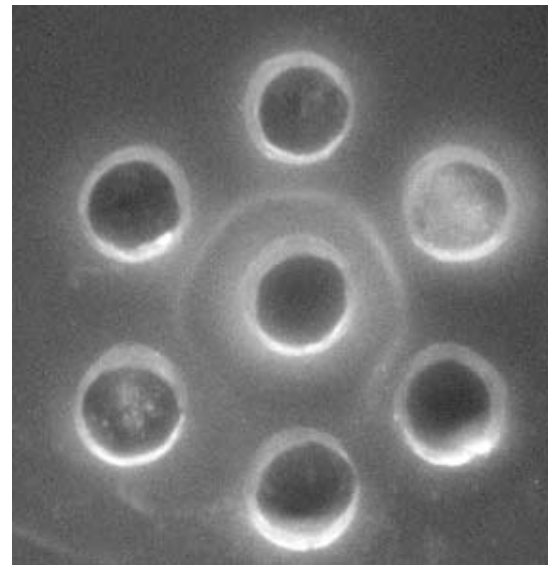
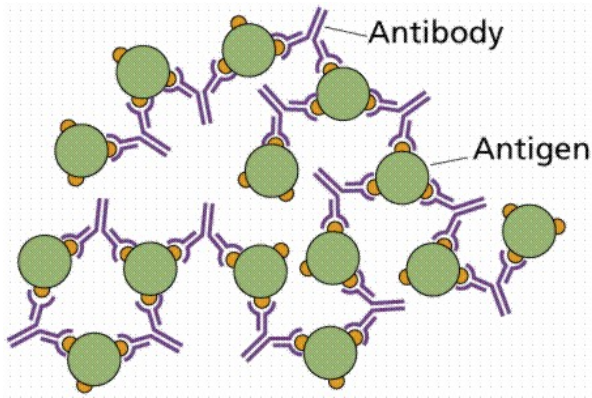
HA-HI

NA-NI

Etc.

# Diagnosis-serology

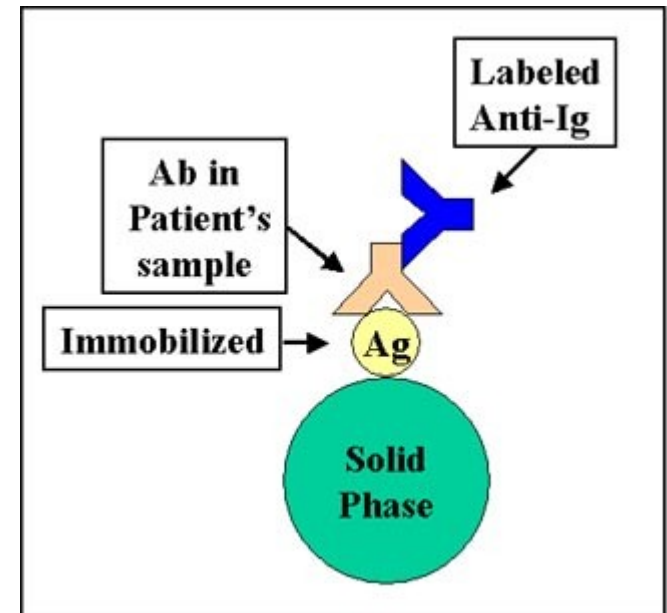
- AGP: Agar Gel Precipitation Test
- Antigen-antibody immune complex precipitate
- migrate through an agar medium
- Type detection



# Diagnosis-serology

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- ELISA: Enzyme-linked immune assay
- Ag+Ab+Enzyme
- AC-EIA: Antigen-capture enzyme immunoassay
- DAS-ELISA: A double-antibody sandwich Elisa
- Dot-Elisa: Dot-enzyme-linked immunosorbent assay
- Type (NP antigen) or subtype (subty antigen) detection



# Diagnosis-serology

VNT: Virus Neutralization Test

a virus-antibody reaction step: the virus is mixed and inoculated with the appropriate antibody

an inoculation step: the mixture of Ag+Ab is inoculated into the host system (e.g. cell cultures, embryonated eggs, or animals)

Detect residual virus infectivity

Ag is known to detect Ab

Ab is known to detect Ag

Detect the subtype

# Diagnosis-serology

- Conventional serological diagnosis is possible by means of the complement fixation and hemagglutination inhibition tests and allows the detection of type and subtype-specific antibodies, respectively.
- As part of an automated serology, immunofluorescence test and enzyme-linked immunosorbent assay are the methods.

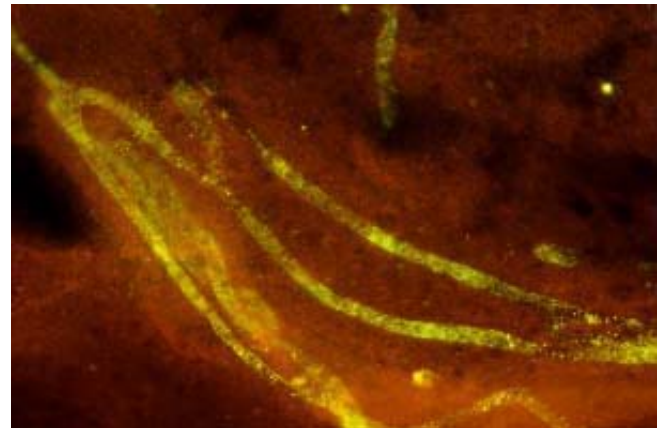
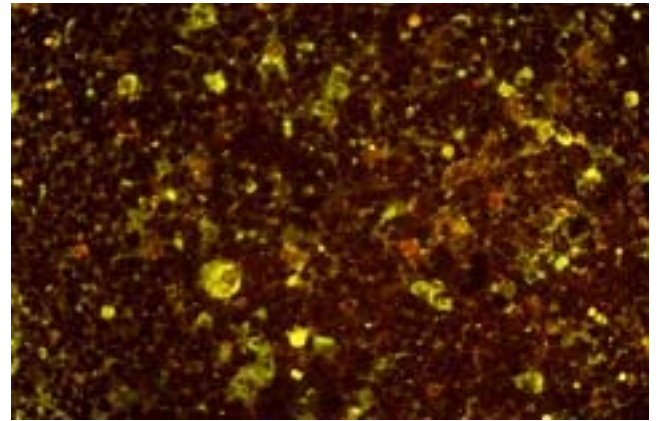
# Diagnosis-serology

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IFA: Immunofluorescence Assay

Ag or Ab labeled by fluorescence, then  
detect the Ag and Ag mixture with  
fluorescence microscope

Type and subtype detection

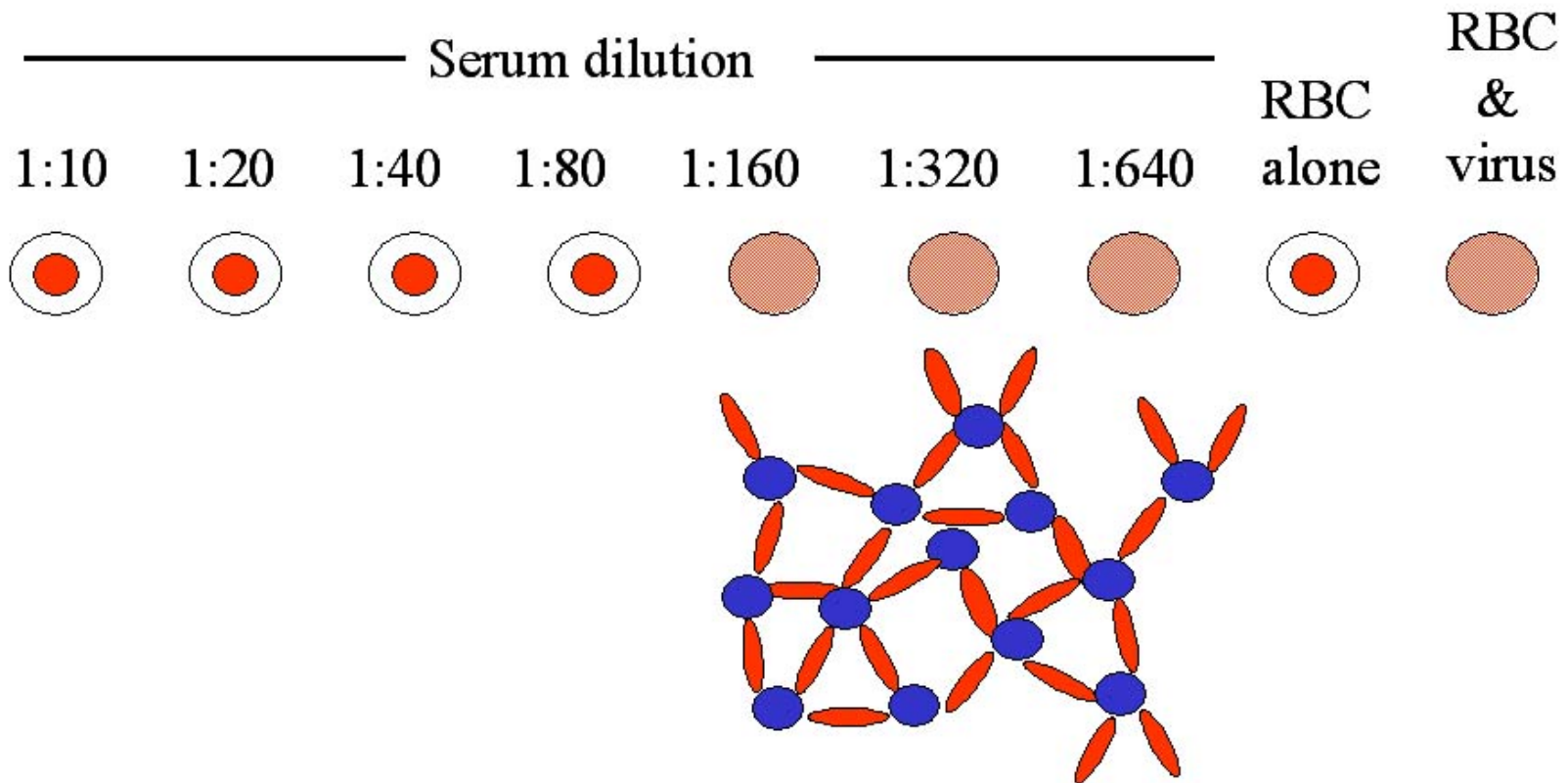




# Diagnosis-serology

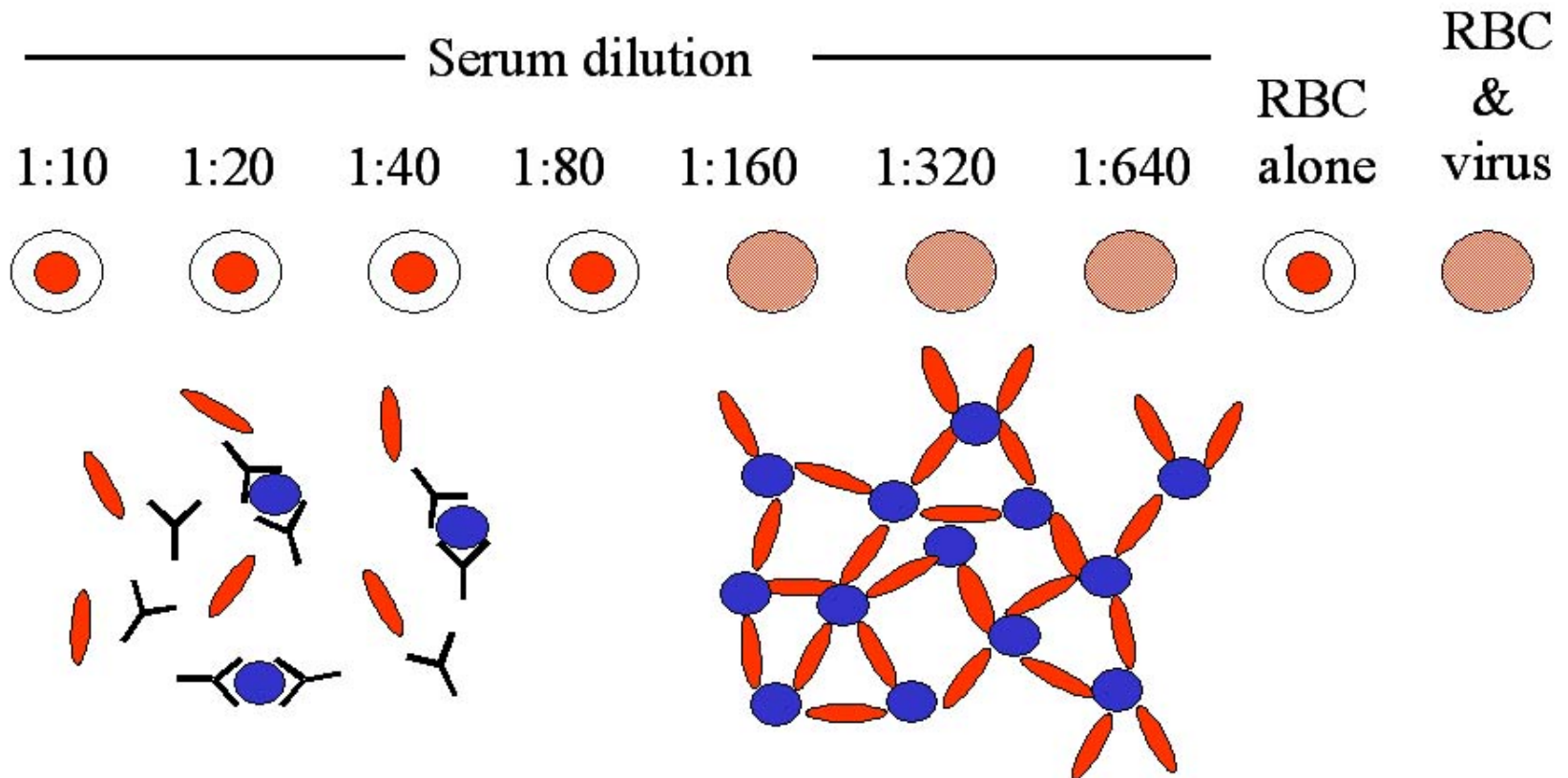
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## Hemagglutination test-HA



# Diagnosis-serology

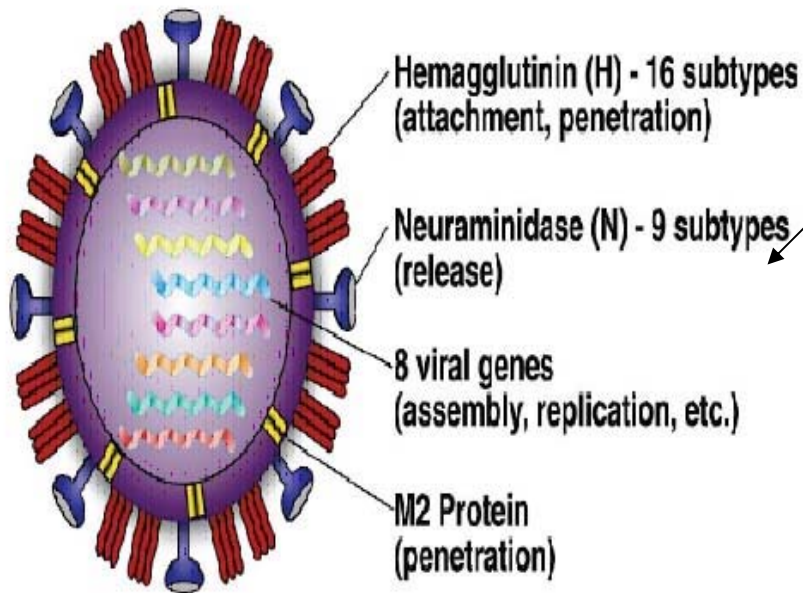
## Hemagglutination Inhibition test-HI



# Diagnosis-serology

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## NA-NI: Neuraminidase Assay and Neuraminidase Inhibition Assay-NA-NI



NA is an enzyme (sialidase) that cleaves terminal sialic acid residues from cell surface receptors of the influenza virus.

Specific attachment of antibody inhibits the activity

# Diagnosis-serology

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NA-NI:



N2(+)

N2 N2

# "Classic" methods for influenza diagnosis

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- Antigen detection using kit-based test on specimens  
- 20 mins
- DFA - 2 to 4 hours
- Virus isolation - 1 to 5 days
- Conventional PCR to sub-type e.g. H5N1 - 1 to 2 days
- Strain determination by sequence e.g. GS/GD/1/96  
- 2 to 7 days
- Need for rapid molecular method

# Differential Diagnosis

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- Virulent Newcastle disease
- Avian pneumovirus
- Infectious laryngotracheitis
- Infectious bronchitis
- Chlamydia
- Mycoplasma
- Acute bacterial diseases
  - Fowl cholera, *E. coli* infection



Thanks for your attention !

